

# Does Impaired Gallbladder Function Contribute to the Development of Barrett's Esophagus and Esophageal Adenocarcinoma?

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## Abstract

**Introduction** Esophageal adenocarcinoma is aetiologically associated with gastro-esophageal reflux, but the mechanisms responsible for the metaplasia–dysplasia sequence are unknown. Bile components are implicated. Impaired gallbladder function may contribute to duodenogastric reflux (DGR) and harmful GERD.

**Aims** This study aims to compare gallbladder function in patients with Barrett's esophagus, adenocarcinoma, and controls.

**Methods** Three groups of patients, all free of gallstone disease, were studied. Group 1: ( $n=15$ ) were normal controls. Group 2: ( $n=15$ ) were patients with >3-cm-long segment of Barrett's esophagus. Group 3: ( $n=15$ ) were patients with esophageal adenocarcinoma. Using real-time ultrasonography unit, gallbladder volume was measured in subjects following a 10-h fast. Ejection fraction was calculated before and after standard liquid meal and compared between the groups.

**Results** The mean percentage reduction in gallbladder volume was 50% at 40 min in the adenocarcinoma group compared with 72.4% in the control group ( $p<0.001$ ). At 60 min, gallbladder filling had recommenced in the control group to 64.1% of fasting volume while continuing to empty with further reduction to 63% in the Barrett's group and to 50.6% ( $p=0.008$ ) in the adenocarcinoma group. The mean gallbladder ejection fraction decreased progressively from controls to Barrett's to adenocarcinoma and was significantly lower in Barrett's group (60.9%;  $p=0.019$ ) and adenocarcinoma group (47.9%;  $p<0.001$ ) compared with normal controls (70.9%).

**Conclusion** Gallbladder function is progressively impaired in Barrett's esophagus and adenocarcinoma. Gallbladder malfunction increases duodenogastric reflux, exposing the lower esophagus to an altered chemical milieu which, in turn, may have a role in promoting metaplasia–dysplasia–neoplasia sequence in the lower esophageal mucosa.

**Keywords** Gallbladder function · Barrett's esophagus · Adenocarcinoma esophagus

## Introduction

The chief risk factor for esophageal adenocarcinoma is Barrett's esophagus, which in turn results from mucosal injury secondary to gastro-esophageal reflux disease (GERD). Impaired lower esophageal sphincter function and the composition of the refluxate are key factors in any resultant mucosal injury. Disturbance of lower esophageal sphincter function allows reflux of gastric contents into the esophagus.<sup>1–4</sup> In addition to gastric acid,<sup>5–7</sup> other components of the refluxate such as pancreatic enzymes,<sup>8–12</sup> intestinal enzymes,<sup>8,13–15</sup> and bile, in particular,<sup>8,16</sup> are all implicated in esophageal mucosal injury.

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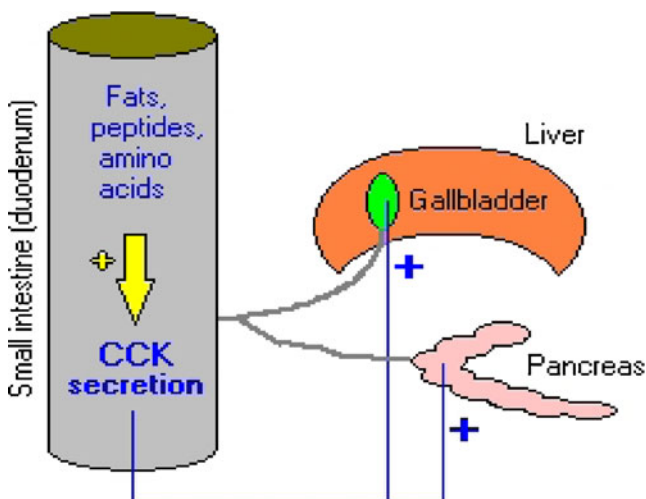
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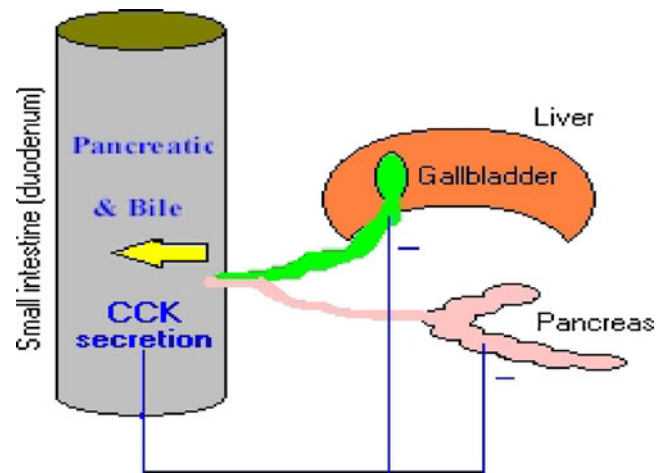
Bile is stored in the gallbladder between meals and is expelled into the duodenum by contraction of the gallbladder in response to meal-stimulated CCK secretion from the duodenum (Fig. 1). In addition to inducing gallbladder contraction, CCK causes relaxation of the sphincter of Oddi and relaxation of the lower esophageal sphincter (LES). The resultant bile bolus in the duodenum switches off CCK release by a negative feedback mechanism. (Fig. 2). Thus, when the gallbladder is functioning normally, bile is mixed with food in the duodenum and little is free to enter the stomach or come into contact with the esophageal mucosa.<sup>18,19</sup>

This orderly sequence of bile processing can be disturbed in a number of circumstances,<sup>20–23</sup> such as a non-functioning gallbladder or following cholecystectomy. When the gallbladder is removed or full of gallstones, the storage facility is destroyed. Instead of storage followed by meal-stimulated release, bile trickles constantly from the liver into the duodenum permitting retrograde reflux into the stomach.<sup>24–26</sup> Furthermore, as the bile is delivered constantly into the duodenum the mechanism for switching off CCK secretion is impaired. Plasma CCK levels remain elevated after meals<sup>27</sup> which in turn may contribute to altered cardia function<sup>28–32</sup> and increased gastro-esophageal reflux. When the gallbladder is non-functioning or poorly functioning bile storage and release by the gallbladder is compromised. The absence of a bolus fails to provide a negative switch-off signal. Gallbladder function may be altered by increasing volumes of gallstones<sup>33,34</sup> chronic cholecystitis, chronic pancreatitis,<sup>35</sup> and by diabetes mellitus<sup>36</sup> amongst other diseases.<sup>37–39</sup>

There is evidence that implicates duodenogastric-esophageal reflux in the pathogenesis of Barrett’s esophagus<sup>40–42</sup> and adenocarcinoma. We have previously shown that cholecystectomy is associated with an increased incidence of GERD.<sup>22,27,70</sup> Others have shown a link between



**Fig. 1** Normal mechanism of CCK release. Meal-stimulated CCK release from the duodenum results in gallbladder emptying. Impaired CCK release is a potential cause of impaired gallbladder emptying



**Fig. 2** Normal mechanism of CCK inhibition. CCK release from the duodenum is inhibited by the negative feedback of a bile bolus in the duodenum. Modulation of CCK release is the chief mechanism of control of gallbladder emptying

cholecystectomy and adenocarcinoma.<sup>20,22,24</sup> Studies have also demonstrated that foregut and gallbladder function are impaired in Barrett’s esophagus.<sup>33,34</sup> There are no studies on gallbladder function in esophageal adenocarcinoma.

As a unifying concept to weave together these strands of evidence, we hypothesized that gallbladder function may be impaired in patients with Barrett’s esophagus and adenocarcinoma of the esophagus, which may in turn contribute to duodenogastric reflux and to the metaplasia–dysplasia–neoplasia sequence. The aim of this study, therefore, was to compare gallbladder function between patients with Barrett’s esophagus, adenocarcinoma, and controls.

## Patients and Methods

### Study Groups

Three study groups were enrolled. Since the prevalence of gallstone disease in patients with Barrett’s is higher than patients without Barrett’s<sup>43</sup>, we screened all patients prior to study to exclude gallstone disease.

- Group 1 (*n*=15) were healthy volunteers attending the radiology department for non-GI radiological investigations. None had symptoms of GERD.
- Group 2 (*n*=15) were patients with histologically confirmed long segment Barrett’s esophagus (>3 cm from the OGJ) who were off all medication that might impact on the gastrointestinal tract for at least 14 days.
- Group 3 (*n*=15) were patients with newly diagnosed adenocarcinoma of the esophagus without evidence of metastatic disease, all of whom were able to swallow fluids without a difficulty.

## Ethics Approval

Approval for the study was obtained from the ethics committee in the hospital before enrolment. All participants were interviewed and a study information proforma was completed. Informed consent was signed by each participant prior to recruitment to the study.

## Exclusion Criteria

Participants with conditions known to affect gallbladder motility were excluded from the study. These included patients with diabetes, chronic liver disease or cholelithiasis. Also excluded were patients being prescribed pharmacological agents known to affect acid secretion or GI or gallbladder motility. A previous history of esophageal, gastric, duodenal, hepatic, pancreatic, or biliary surgery was also an exclusion factor. Esophageal adenocarcinoma patients with advanced cancer, an esophageal stent in situ, severe dysphagia that might compromise the ingestion of the test meal, or patients receiving chemoradiotherapy or post-chemoradiotherapy were also excluded. Apart from the adenocarcinoma group none had a history of cancer.

## Sample Size

Sample size was calculated using “PS: Power and Sample Size Calculation<sup>©</sup>” software (version 2.1.31, 2003, Department of Biostatistics, Vanderbilt University School of Medicine, Nashville, TN).

## The Standard Test Meal

All patients were given a standard test liquid meal to stimulate gallbladder contraction. The meal used as the standard test meal was the commercially available Fortisip<sup>®</sup> Strawberry (Nutricia Clinical NZ, Auckland, New Zealand). Fortisip<sup>®</sup> Strawberry is a commercially available nutritionally complete food commonly used for patients with increased energy or protein requirements or for those who have little appetite for food. A liquid preparation was chosen as standard to facilitate rapid ingestion into the stomach for patients with some degree of stenosis. The meal provided 300 kcal in each 200 ml fluid preparation (1.5 kcal/ml) in the form of 10.2% saturated fatty acids, 9.4% monounsaturated fatty acids, and 30.4% polyunsaturated fatty acids. It was felt that this would facilitate gallbladder emptying in a more physiological manner than using a purely fatty meal which may have resulted in exaggerated emptying. The standard test meal (Fortisip) was ingested at a rate of 50–100 ml/min via a straw.<sup>44,45</sup>

## Measurement of Gallbladder Function

A real-time ultrasonography unit (Model: GE Logiq 9, GE Healthcare Technologies, Clinical Systems Information Technologies, Hatfield, UK) was used for measuring gallbladder volume.<sup>46</sup> The transducer used was a real-time multi-frequency (2–4 MHz) sector transducer.

Ultrasonographic scanning was performed after a 10-h overnight fast to ensure adequate gallbladder distension. The fasting gallbladder volume (FGV) was calculated and the common bile duct caliber was measured before meal ingestion. The gallbladder volume (GBV) was also calculated at 20, 40, and 60 min following the standard test meal.

Study participants remained in the sitting position after meal and between scans to facilitate passage of the meal through the stomach to the duodenum. Any participant who was unable to tolerate sitting comfortably adopted the right lateral position. Images and measurements were obtained in suspended deep inspiration.

Gallbladder volume was calculated using software capable of automatically capturing the gallbladder length, width and depth (Model: GE Logiq 9, GE Healthcare Technologies, Clinical Systems Information Technologies, Hatfield, UK). To verify the software produced readings, duplicate readings were also obtained to calculate the gallbladder volume manually by using the ellipsoid formula:<sup>39,46,47</sup>

$$\text{Volume} = \frac{\pi \times \text{Length} \times \text{Width} \times \text{Depth}}{6}$$

The FGV was taken as the base line volume.

## Calculation of Gallbladder Emptying

Gallbladder emptying (GBE) was taken as the difference between the FGV and the GBV at a specific time, expressed as a percentage of the basal gall bladder volume:

$$\text{GBE}(X_{\min}) = \frac{(\text{FGV} - \text{GBV}(X_{\min})) \times 100}{(\text{FGV})}$$

## Data Collection and Statistical Analysis

Data were collected using a computer-generated database (Microsoft Office Access 2003, Microsoft Windows XP Professional<sup>™</sup>, Microsoft Corporation, Redmond, WA). Statistical analysis was performed using SPSS 10.0 software for Windows<sup>™</sup> (SPSS Inc., Chicago, IL) to calculate the mean, the standard deviation (SD) and the any significant difference between the different groups. The *t* test for independent samples was used to compare gallbladder emptying and ejection fraction between the study groups and the control group using a *p* value of <0.05 and a power of 0.8.

## Results

### Demographic Data

A total of forty-five participants had gallbladder emptying assessed with a male/female ratio of 2.4:1. There was no significant difference between the ages of the different groups. The mean (SD) age of the control group was 69.7 (8.7) years which compared with 66.0 (9.9) years for the Barrett's group ( $p=0.542$ ) and 64.5 (14.9) years for the adenocarcinoma group ( $p=0.622$ ).

### Gallbladder Volume

The mean resting gallbladder volume was 26(1.1) mls in controls, compared with 38.1 (26.8) ml in the Barrett's group ( $p=0.005$ ) and 27.6(16.1) ml in the adenocarcinoma group ( $p=0.054$ ) (Table 1).

Gallbladder volume decreased gradually after the standard meal in all groups. This decrease was more significant at 20-min posttest meal in the control group, where gallbladder volume fell to 10.5 (5.1) ml than in the Barrett's group 22.7 (21.2) ml compared with the control group ( $p<0.001$ ), or the adenocarcinoma group 15.5 (15.7) ml compared with the control group ( $p=0.003$ ).

Gallbladder volume reached its lowest level in the control group at 40 min to 7.5<sup>4</sup> ml and had started to fill again by 60 min to 9.4 (4.3) ml. Both the Barrett's and the adenocarcinoma groups continued to show significant decrease in volume at 60 min confirming continuation of emptying with volumes of 14.9 (14.5) ml ( $p=0.007$ ) and 14.4<sup>13</sup> ml ( $p=0.01$ ), respectively. The gallbladder volume decrease was slower in the Barrett's group and the cancer group than in normal controls and continued up to 60 min with no recovery.

### Percentage of Gallbladder Emptying

Because the mean starting fasting gallbladder volume showed slight variability among the three groups, the

percentage gallbladder emptying was used to provide better comparison between the three study groups. The fasting gallbladder volume was taken as the base line volume from which subsequent emptied volumes were calculated as the percentage of gallbladder emptying at a specific point of time.

The mean percentage gallbladder volume emptied at 20 min following a standard meal in the control group was 60.5% (12.5%) which was not significantly different from the 48.5% (18%) in the Barrett's group. The percentage emptying in the adenocarcinoma group was 41% (26%) which was significantly reduced compared with the control group ( $p=0.021$ ).

At 40 min, the mean percentage gallbladder volume reduction in the control group was 72.4% (6.9%), which was similar to the Barrett's group 58.4% (11.6%;  $p=0.213$ ) but was significantly different from the adenocarcinoma group 44.9% (20.7;  $p<0.001$ ).

At 60 min, gallbladder filling had recommenced in the control group to 64.1% (10.2). Both the Barrett's and the adenocarcinoma groups continued to show progression of emptying with further reduction to 63%<sup>13</sup> ( $p=0.427$ ) in the Barrett's group and 50.6% (26.5;  $p=0.008$ ) in the adenocarcinoma group.

The gallbladder emptying was faster and more significant in the control group compared with the cancer group but not the Barrett's group, mirroring the volume changes, and reaching maximal emptying at 40 min after which recovery was seen. Gallbladder emptying was delayed in the Barrett's and the cancer group particularly as seen after 40 min and continued up to 60 min posttest meal.

### Gallbladder Ejection Fraction

The gallbladder ejection fraction was taken as gallbladder emptying at 40 min. The ejection fraction was taken as the study's end-point at which the statistical test for the study power was taken. The ejection fraction was calculated as 72.4% (6.9) in the control group, 58.4% (11.6) in the Barrett's group ( $p=0.213$ ), and 44.9% (20.7) in the adenocarcinoma group ( $p<0.001$ ) using the Student's *t* test for independent samples.

**Table 1** Gallbladder volumes

	Fasting GBV (ml)	20 min (ml)	40 min (ml)	60 min (ml)
Controls	26	10.5	8	9.4
Barrett's group	38.1	22.7	15	15
Adenocarcinoma	27.6	15.5	18	14.4

Gallbladder volume decreased gradually after the standard meal in all groups. The gallbladder volume decrease was slower in the Barrett's group and the cancer group and continued up to 60 min with no recovery

## Discussion

The increase in the incidence esophageal adenocarcinoma is multifactorial, but the contribution of Barrett's esophagus is beyond question.<sup>48</sup> The aetiology of Barrett's mucosa, which represents the severest end of the reflux spectrum, is in turn a consequence of altered gastrointestinal anatomy and physiological mechanisms<sup>49</sup> the most significant of which are the alteration in intestinal motility<sup>50</sup> and the



toxicity of the resultant refluxate.<sup>16,51,52</sup> The motility disturbances identified in Barrett's include esophageal body dysmotility leading to impaired clearance<sup>53,54</sup> lower mean basal LES pressure and increased transient lower esophageal sphincter relaxation episodes, which allow increased gastro-esophageal reflux, gastric dysmotility leading to delayed gastric emptying which could promote reflux and prolong contact with toxic refluxate<sup>55–60</sup> and possible antro-duodenal motility disorders.<sup>61,62</sup>

The toxicity of the refluxate is clearly a co-contributing factor to the resultant injury. While we are unclear about the exact chemical components which inflict most injury, it is clear that injury may result from gastric acid alone<sup>52</sup> or from acid combined with duodenal refluxate.<sup>63,64</sup> The role of acid and bile in the genesis of esophageal mucosal damage and reflux symptoms is complex. Acid combined with pepsin and unconjugated bile acids are critical to the development of esophagitis and Barrett's esophagus.<sup>40,42</sup> Bile alone or duodenal refluxate alone may be the principal factor in determining the severity of esophagitis as Barrett's has been described following total gastrectomy.<sup>65,66</sup> Patients with reflux disease have an increased concentration of bile acids in their esophageal aspirates.<sup>12</sup> Nearly half of the patients with reflux symptoms have combined pathological acid and bile reflux.<sup>67</sup>

Duodenogastric reflux (DGR) of duodenal content into the stomach occurs physiologically but anything that alters the structure or function of the duodeno-pancreatobiliary system will promote DGR. Thus, surgical destruction of the pylorus after distal gastrectomy,<sup>68</sup> pyloroplasty or Whipples procedure increase DGR.<sup>17</sup> Similarly, anything that disturbs the orderly sequence of bile secretion, storage, and release may contribute to DGR.<sup>69</sup> The most dramatic alteration in this environment occurs after cholecystectomy when bile is no longer stored between meals but streams continuously into the duodenum and the stomach.<sup>22,23</sup> We, and others, have previously shown that cholecystectomy results in increased gastro-esophageal reflux<sup>70</sup> and elevated levels of CCK.<sup>27</sup> Amongst the effects of CCK is the reduction in LOS pressure.<sup>27</sup> These combined disturbances may contribute to an abnormal concentration of bile refluxate for a longer duration in the lower esophagus. These findings support a possible contribution of gallbladder malfunction in the development of Barrett's esophagus and adenocarcinoma.<sup>21</sup>

In this study we have shown that gallbladder emptying is progressively impaired in the Barrett's esophagus and adenocarcinoma groups. In normal subjects gallbladder emptying in response to a meal was complete by 40 min after which it began to fill again. Approximately two thirds of resting volume had emptied within the first 20 min and three quarters by 40 min and the filling process had commenced within 60 min of a meal, which is consistent with the literature.<sup>71</sup> The pattern of emptying was altered in

the Barrett's group and even more so in the adenocarcinoma group where both groups continued to show significant decrease in volume to 60 min confirming continuation of emptying suggesting abnormal gallbladder motility. Thus, patients with Barrett's and adenocarcinoma had an impaired response to meals suggesting that their gallbladder function was impaired and incapable of storing and releasing bile normally, a condition approaching non-functioning gallbladder. Patient following cholecystectomy<sup>21–23</sup> or patients with poorly functioning gallbladders have increased bile reflux into the stomach and increased potential toxicity.

The cause of the abnormal gallbladder function in Barrett's and cancer is unclear. It may be that there is a common motility disorder that predisposes to both Barrett's esophagus and gallbladder malfunction. This may have a neural basis or a hormonal basis, most likely through CCK secretion. Whatever the cause, it is likely that a lifetime of altered gallbladder function may predispose to chronic toxic bile exposure in the stomach and lower esophagus. Barrett's esophagus is associated with motility disorders of the esophagus and of the stomach and a recent report has observed an association between Barrett's and gallbladder function.<sup>43</sup> Our study is the first report to describe an association between gallbladder malfunction and adenocarcinoma. It is possible that there is a primary motility disorder in Barrett's esophagus affecting the entire upper gastrointestinal tract affecting the esophagus, stomach and the biliary system.

In conclusion, patients with Barrett's esophagus and esophageal adenocarcinoma have abnormal gallbladder function. Whether this is cause or consequence is unclear. Further studies are needed to determine whether impaired gallbladder function contributes significantly to the reflux milieu and in particular to the mucosal change of Barrett's esophagus or adenocarcinoma.

## References

1. Gillen P, Thornton J, Byrne PJ, Walsh TN, Hennessy TP. Implications of upright gastro-oesophageal reflux. *Br J Surg* 1994;81(2):239–40.
2. Holland CT, Satchell PM, Farrow BR. Vagal afferent dysfunction in naturally occurring canine esophageal motility disorder. *Dig Dis Sci* 1994;39(10):2090–8.
3. Klauser AG, Voderholzer WA, Knesewitsch PA, Schindlbeck NE, Muller-Lissner SA. What is behind dyspepsia? *Dig Dis Sci* 1993;38(1):147–54.
4. Lidums I, Holloway R. Motility abnormalities in the columnar-lined esophagus. *Gastroenterol Clin North Am* 1997;26(3):519–31.
5. Bruley des Varannes S, Ravenbakht-Charifi M, Cloarec D, Pujol P, Simon J, Galmiche JP. [Barret's esophagus and acid gastroesophageal reflux. Two-channel pH-metric measurements and manometric study]. *Gastroenterol Clin Biol* 1992;16(5):406–12.

6. Champion G, Richter JE, Vaezi MF, Singh S, Alexander R. Duodenogastric reflux: relationship to pH and importance in Barrett's esophagus. *Gastroenterology* 1994;107(3):747–54.
7. Mittal RK, Lange RC, McCallum RW. Identification and mechanism of delayed esophageal acid clearance in subjects with hiatus hernia. *Gastroenterology* 1987;92(1):130–5.
8. Stipa F, Stein HJ, Feussner H, Kraemer S, Siewert JR. Assessment of non-acid esophageal reflux: comparison between long-term reflux aspiration test and fiberoptic bilirubin monitoring. *Dis Esophagus* 1997;10(1):24–8.
9. Yamashita Y, Homma K, Kako N, Clark GW, Smyrk TC, Hinder RA, et al. Effect of duodenal components of the refluxate on development of esophageal neoplasia in rats. *J Gastrointest Surg* 1998;2(4):350–5.
10. Evander A, Little AG, Riddell RH, Walther B, Skinner DB. Composition of the refluxed material determines the degree of reflux esophagitis in the dog. *Gastroenterology* 1987;93(2):280–6.
11. Mud HJ, Kranendonk SE, Obertop H, Van Houten H, Westbroek DL. Active trypsin and reflux oesophagitis: an experimental study in rats. *Br J Surg* 1982;69(5):269–72.
12. Imada T, Chen C, Hatori S, Shiozawa M, Rino Y. Effect of trypsin inhibitor on reflux oesophagitis after total gastrectomy in rats. *Eur J Surg* 1999;165(11):1045–50.
13. Castell DO, Murray JA, Tutuian R, Orlando RC, Arnold R. Review article: the pathophysiology of gastro-oesophageal reflux disease - oesophageal manifestations. *Aliment Pharmacol Ther* 2004;20 Suppl 9:14–25.
14. Naito Y, Uchiyama K, Kuroda M, Takagi T, Kokura S, Yoshida N, et al. Role of pancreatic trypsin in chronic esophagitis induced by gastroduodenal reflux in rats. *J Gastroenterol* 2006;41(3):198–208.
15. Lillemoen KD, Johnson LF, Harmon JW. Alkaline esophagitis: a comparison of the ability of components of gastroduodenal contents to injure the rabbit esophagus. *Gastroenterology* 1983;85(3):621–8.
16. Nehra D, Howell P, Williams CP, Pye JK, Beynon J. Toxic bile acids in gastro-oesophageal reflux disease: influence of gastric acidity. *Gut* 1999;44(5):598–602.
17. Kuran S, Parlak E, Aydog G, Kacar S, Sasmaz N, Ozden A, et al. Bile reflux index after therapeutic biliary procedures. *BMC Gastroenterol*. 2008;8:4.
18. Koop I. Role of bile acids in the control of pancreatic secretion and CCK release. *Eur J Clin Invest* 1990;20 Suppl 1:S51–7.
19. Szanto I, Voros A, Gonda G, Nagy P, Cserepes E, Gamal EM, et al. [Esophageal implantation metastasis from adenocarcinoma of the cardia]. *Magy Seb* 2001;54(6):393–6.
20. Manifold DK, Anggiansah A, Owen WJ. Effect of cholecystectomy on gastroesophageal and duodenogastric reflux. *Am J Gastroenterol* 2000;95(10):2746–50.
21. Freedman J, Ye W, Naslund E, Lagergren J. Association between cholecystectomy and adenocarcinoma of the esophagus. *Gastroenterology* 2001;121(3):548–53.
22. Jazrawi S, Walsh TN, Byrne PJ, Hennessy TP. Cholecystectomy and oesophageal pathology: is there a link? *Ir J Med Sci* 1993;162(6):209–12.
23. Soran A, Erverdi N, Col C, Aslar K, Cete M, Hengrimen S (2000). "Effect of Cholecystectomy on Duodenogastric Reflux, Gastric Mucosa and Serum Gastrin Level." *Nagoya Medical Journal* 44(1): 19–25.
24. Cabrol J, Navarro X, Simo-Deu J, Segura R. Evaluation of duodenogastric reflux in gallstone disease before and after simple cholecystectomy. *Am J Surg* 1990;160(3):283–6.
25. Brough WA, Taylor TV, Torrance HB. The surgical factors influencing duodenogastric reflux. *Br J Surg* 1984;71(10):770–3.
26. Brough WA, Taylor TV, Torrance HB. The effect of cholecystectomy on duodenogastric reflux in patients with previous peptic ulcer surgery. *Scand J Gastroenterol Suppl* 1984;92:255–6.
27. McDonnell CO, Bailey I, Stumpf T, Walsh TN, Johnson CD. The effect of cholecystectomy on plasma cholecystokinin. *Am J Gastroenterol* 2002;97(9):2189–92.
28. Wilson P, Welch NT, Hinder RA, Anselmino M, Herrington MK, DeMeester TR, et al. Abnormal plasma gut hormones in pathologic duodenogastric reflux and their response to surgery. *Am J Surg* 1993;165(1):169–76; discussion 176–7.
29. Boulant J, Mathieu S, D'Amato M, Abergel A, Dapoigny M, Bommelaer G. Cholecystokinin in transient lower oesophageal sphincter relaxation due to gastric distension in humans. *Gut* 1997;40(5):575–81.
30. Zerbib F, Bruley Des Varannes S, Scarpignato C, Leray V, D'Amato M, Roze C, et al. Endogenous cholecystokinin in postprandial lower esophageal sphincter function and fundic tone in humans. *Am J Physiol* 1998;275(6 Pt 1):G1266–73.
31. Clave P, Gonzalez A, Moreno A, Lopez R, Farre A, Cusso X, et al. Endogenous cholecystokinin enhances postprandial gastroesophageal reflux in humans through extrasphincteric receptors. *Gastroenterology* 1998;115(3):597–604.
32. Ledebor M, Masclee AA, Batstra MR, Jansen JB, Lamers CB. Effect of cholecystokinin on lower oesophageal sphincter pressure and transient lower oesophageal sphincter relaxations in humans. *Gut* 1995;36(1):39–44.
33. Portincasa P, Di Ciaula A, Palmieri V, Velardi A, VanBerge-Henegouwen GP, Palasciano G. Impaired gallbladder and gastric motility and pathological gastro-oesophageal reflux in gallstone patients. *Eur J Clin Invest* 1997;27(8):653–61.
34. O'Donnell LJ, Fairclough PD. Gall stones and gall bladder motility. *Gut* 1993;34(4):440–3.
35. Meguro T, Shimosegawa T, Kashimura J, Kikuchi Y, Koizumi M, Toyota T. Gallbladder emptying to endogenous and exogenous stimulation in chronic pancreatitis patients. *Am J Gastroenterol* 1994;89(2):225–31.
36. Shaw SJ, Hajnal F, Lebovitz Y, Ralls P, Bauer M, Valenzuela J, et al. Gallbladder dysfunction in diabetes mellitus. *Dig Dis Sci* 1993;38(3):490–6.
37. Jutras JA. Hyperplastic cholecystoses; Hickey lecture, 1960. *Am J Roentgenol Radium Ther Nucl Med* 1960;83:795–827.
38. Swayne LC, Heitner D, Rubenstein JB, Fernandez A, Niknejad G. Differential gallbladder contractility in fundal adenomyomatosis: demonstration by cholecystokinin cholescintigraphy. *J Nucl Med* 1987;28(11):1771–4.
39. Acalovschi M, Dumitrascu DL, Nicoara CD. Gallbladder contractility in liver cirrhosis: comparative study in patients with and without gallbladder stones. *Dig Dis Sci* 2004;49(1):17–24.
40. Souza RF, Krishnan K, Spechler SJ. Acid, bile, and CDX: the ABCs of making Barrett's metaplasia. *Am J Physiol Gastrointest Liver Physiol*. 2008 Aug;295(2):G211–8.
41. Gutschow CA, Bludau M, Vallbohmer D, Schroder W, Bollsweiler E, Holscher AH. NERD, GERD, and Barrett's esophagus: role of acid and non-acid reflux revisited with combined pH-impedance monitoring. *Dig Dis Sci*. 2008 Dec;53(12):3076–81.
42. Triadafilopoulos G. Acid and bile reflux in Barrett's esophagus: a tale of two evils. *Gastroenterology*. 2001 Dec;121(6):1502–6.
43. Izbeki F RA, Yobuta JS, Roka R, Lonovics J, Wittmann T. Increased prevalence of gallstone disease and impaired gallbladder motility in patients with Barrett's esophagus. *Dig Dis Sci*. 2008(53):2268–75.
44. Bobba VR, Krishnamurthy GT, Kingston E, Turner FE, Brown PH, Langrell K. Gallbladder dynamics induced by a fatty meal in normal subjects and patients with gallstones: concise communication. *J Nucl Med* 1984;25(1):21–4.
45. Dodds WJ, Groh WJ, Darweesh RM, Lawson TL, Kishk SM, Kern MK. Sonographic measurement of gallbladder volume. *AJR Am J Roentgenol* 1985;145(5):1009–11.
46. Kishk SM, Darweesh RM, Dodds WJ, Lawson TL, Stewart ET, Kern MK, et al. Sonographic evaluation of resting gallbladder

- volume and postprandial emptying in patients with gallstones. *AJR Am J Roentgenol* 1987;148(5):875–9.
47. Acalovschi M, Dumitrascu DL, Suteu T, Veres A, Albu S, Badea RI. Misoprostol induces gallbladder contraction during fasting, but does not influence postprandial emptying: an ultrasound study in healthy subjects. *Acta Gastroenterol Belg* 2002;65(4):191–5.
  48. Dickman R, Kim JL, Camargo L, Green SB, Sampliner RE, Garewal HS, Fass R: Correlation of gastroesophageal reflux disease symptoms characteristics with long-segment Barrett's esophagus. *Dis Esophagus* 2006;19:360–365
  49. Dent J: From 1906 to 2006—a century of major evolution of understanding gastroesophageal reflux disease. *Aliment Pharmacol Ther* 2006;24:1269–1281.
  50. Ang D, Blondeau K, Sifrim D, Tack J. The spectrum of motor function abnormalities in gastroesophageal reflux disease and Barrett's esophagus. *Digestion*. 2009;79(3):158, 68.
  51. Kauer WK, Peters JH, DeMeester TR, Ireland AP, Bremner CG, Hagen JA. Mixed reflux of gastric and duodenal juices is more harmful to the esophagus than gastric juice alone. The need for surgical therapy re-emphasized. *Ann Surg*. 1995 Oct; 222(4):525–31; discussion 31–3.
  52. Richter JE. Role of the gastric refluxate in gastroesophageal reflux disease: acid, weak acid and bile. *Am J Med Sci*. 2009 Aug;338(2):89–95.
  53. Parkman HP, Fisher RS: Contributing role of motility abnormalities in the pathogenesis of gastroesophageal reflux disease. *Dig Dis* 1997;15:40–52
  54. Holloway RH: Esophageal body motor response to reflux events: secondary peristalsis. *Am J Med* 2000;108(suppl 4A):20S–26S.
  55. Byrne PJ, Mulligan ED, O'Riordan J, Keeling PWN, Reynolds JV: Impaired visceral sensitivity to acid reflux in patients with Barrett's esophagus. The role of esophageal motility. *Dis Esophagus* 2003;16:199–203.
  56. McCallum RW, Berkowitz DM, Lerner E: Gastric emptying in patients with gastroesophageal reflux. *Gastroenterology* 1981;80:285–291.
  57. Maddern GJ, Chatterton BE, Collins PJ, Horowitz M, Shearman DJC, Jamieson GG: Solid and liquid emptying in patients with gastroesophageal reflux. *Br J Surg* 1985;72:344–347.
  58. Cunningham KM, Horowitz M, Riddel PS: Relations among autonomic nerve dysfunction, esophageal motility and gastric emptying in gastroesophageal reflux disease. *Gut* 1991;32:1436–1440.
  59. Benini L, Sembenini C, Castellani G, Caliari S, Fioretta A, Vantini I: Gastric emptying and dyspeptic symptoms in patients with gastroesophageal reflux. *Am J Gastroenterol* 1996;91:1351–1354.
  60. Buckles D, Sarosiek I, McMillin C, McCallum R: Delayed gastric emptying in gastroesophageal reflux disease: reassessment with new methods and symptomatic correlations. *Am J Med Sci* 2004;327:1–4.
  61. Parkman HP, Harris AD, Krevsky B, Urbain JL, Maurer AH, Fisher RS: Gastrointestinal motility and dysmotility: an update on techniques available for evaluation. *Am J Gastroenterol* 1995;90:869–892.
  62. Vaezi MF, Richter GE: Role of acid and duodenogastroesophageal reflux in gastroesophageal reflux disease. *Gastroenterology* 1996;111:1192–1199
  63. Schweitzer EJ, Bass BL, Batzri S, Harmon JW. Bile acid accumulation by rabbit esophageal mucosa. *Dig Dis Sci* 1986;31(10):1105–13.
  64. Schweitzer EJ, Harmon JW, Bass BL, Batzri S. Bile acid efflux precedes mucosal barrier disruption in the rabbit esophagus. *Am J Physiol* 1984;247(5 Pt 1):G480–5.
  65. Sinn DH, Kim KM, Kim ER, Son HJ, Kim JJ, Rhee JC, Development of Barrett's. Esophagus soon after total Gastrectomy. *Gut and Liver* 2008; 2:51–53.
  66. Yuasa N, Abe T, Sasaki E, Fukaya M, Nimura Y, Miyahara R, Comparison of gastroesophageal reflux in 100 patients with or without prior gastroesophageal surgery. *J Gastroenterology*. 2009; 44(7): 650–658.
  67. Fein M, Maroske J, Fuchs KH. Importance of duodenogastric reflux in gastro-oesophageal reflux disease. *Br J Surg*. 2006 Dec;93(12):1475–82.
  68. Ritchie WP: Alkaline reflux gastritis. *Gastroenterol Clin North Am* 1994;23:281–294
  69. Malekzadeh R, Nasser-Moghaddam S, Sotoudeh M, Gastroesophageal reflux disease: The New Epidemic, *Arch Iranian Med*,2003; 6: 127–140.
  70. Jazrawi S, Walsh TN, Byrne PJ, Hill AD, Li H, Lawlor P, et al. Cholecystectomy and oesophageal reflux: a prospective evaluation. *Br J Surg* 1993;80(1):50–3.
  71. Katz PO. Review article: the role of non-acid reflux in gastro-oesophageal reflux disease. *Aliment Pharmacol Ther* 2000;14(12):1539–51.